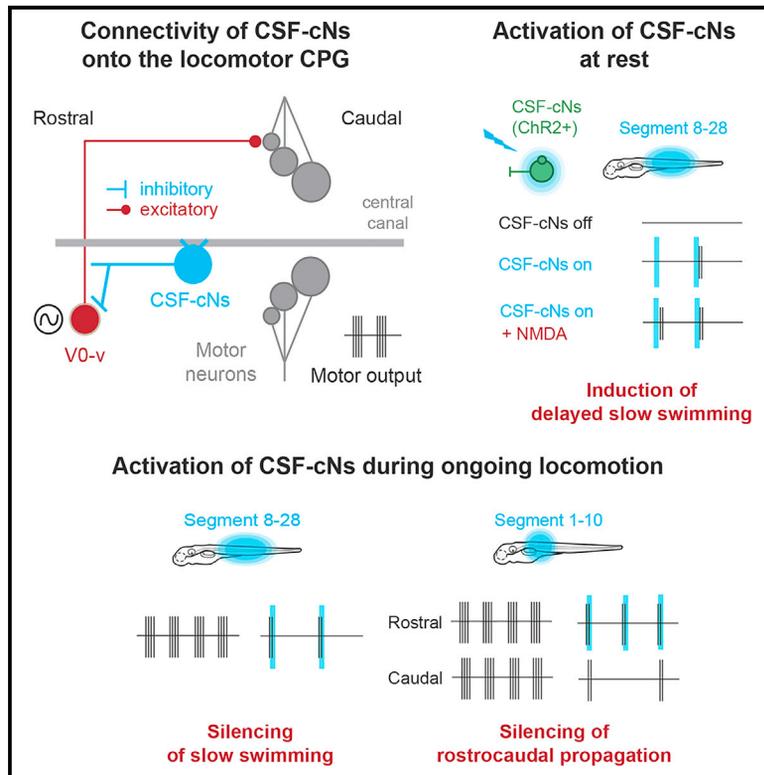


State-Dependent Modulation of Locomotion by GABAergic Spinal Sensory Neurons

Graphical Abstract



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In Brief

Fidelin et al. demonstrate how GABAergic spinal sensory neurons modulate circuits underlying slow locomotion in the zebrafish larva in a state-dependent manner. CSF-cNs gate the rostrocaudal propagation of excitation and modulate duration and frequency of locomotor events by contacting essential premotor interneurons of the slow swimming CPG.

Highlights

- Spinal CSF-cNs project onto glutamatergic premotor interneurons
- CSF-cNs exert a state-dependent modulation of spinal premotor circuits
- CSF-cNs modulate duration and frequency of locomotor events
- CSF-cNs gate rostrocaudal propagation of excitation in the spinal cord



State-Dependent Modulation of Locomotion by GABAergic Spinal Sensory Neurons

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SUMMARY

The cerebrospinal fluid (CSF) constitutes an interface through which chemical cues can reach and modulate the activity of neurons located at the epithelial boundary within the entire nervous system. Here, we investigate the role and functional connectivity of a class of GABAergic sensory neurons contacting the CSF in the vertebrate spinal cord and referred to as CSF-cNs. The remote activation of CSF-cNs was shown to trigger delayed slow locomotion in the zebrafish larva, suggesting that these cells modulate components of locomotor central pattern generators (CPGs). Combining anatomy, electrophysiology, and optogenetics *in vivo*, we show that CSF-cNs form active GABAergic synapses onto V0-v glutamatergic interneurons, an essential component of locomotor CPGs. We confirmed that activating CSF-cNs at rest induced delayed slow locomotion in the fictive preparation. In contrast, the activation of CSF-cNs promptly inhibited ongoing slow locomotion. Moreover, selective activation of rostral CSF-cNs during ongoing activity disrupted rostrocaudal propagation of descending excitation along the spinal cord, indicating that CSF-cNs primarily act at the premotor level. Altogether, our results demonstrate how a spinal GABAergic sensory neuron can tune the excitability of locomotor CPGs in a state-dependent manner by projecting onto essential components of the excitatory premotor pool.

INTRODUCTION

During active locomotion, sensory afferent neurons provide excitatory feedback to motor neurons and spinal interneurons in response to muscle contraction. Local GABAergic interneurons can modulate this pathway by inhibiting sensory afferents at the presynaptic level [1, 2]. Genetic targeting and manipulation of these GABAergic interneurons recently demonstrated

the importance of presynaptic modulation of sensory afferents to control fine motor behaviors in mice [3, 4]. Although GABAergic modulation is essential for controlling excitability and spike timing of excitatory neurons throughout the nervous system [5, 6], little is known about the GABAergic modulation of descending premotor excitatory interneurons controlling rhythm and pattern generation in the spinal cord. Pharmacological manipulations showed that GABAergic neurons could modulate the burst frequency of motor neurons during fictive locomotion [7, 8], suggesting that the release of GABA can control the excitability of spinal excitatory interneurons driving fictive locomotion. Yet, the nature of GABAergic neurons mediating this effect and their targets in the spinal cord remain to be identified.

Almost a century ago, Kolmer and Agduhr identified cerebrospinal fluid-contacting neurons (CSF-cNs), also called KA cells in African clawed frog and zebrafish, as villiated neurons surrounding the central canal in the spinal cord of over 200 vertebrate species [9–11]. Spinal CSF-cNs are sensory neurons with unique features because they are GABAergic, intraspinal, and reside in the ventral part of the spinal cord. As CSF-cNs exhibit longitudinal axons projecting in the ventral cord, they might relay chemical information from the cerebrospinal fluid (CSF) to spinal circuits [12–14]. Despite the conservation of CSF-cNs among vertebrates, their role in sensorimotor integration is still poorly understood. Remote activation of CSF-cNs was shown to trigger delayed slow locomotor activity in head-restrained zebrafish larvae, indicating that CSF-cNs could project onto components of the slow swimming central pattern generator (CPG) [14]. However, the nature of postsynaptic targets of CSF-cNs within the slow locomotor CPG remains to be identified.

Here, we took advantage of the zebrafish larva to optically probe the cellular and circuit mechanisms deployed by GABAergic CSF-cNs neurons to modulate locomotor activity in an intact animal. We generated a specific line to analyze the morphology of CSF-cNs, manipulate their activity, and map their functional connectivity onto locomotor CPGs. We performed channelrhodopsin-2 (ChR2)-mediated activation of CSF-cNs in combination with whole-cell recordings of their targets to demonstrate that CSF-cNs form GABAergic synapses onto *dbx1⁺/evx1⁺* glutamatergic commissural descending V0-v interneurons. These interneurons are essential components of the

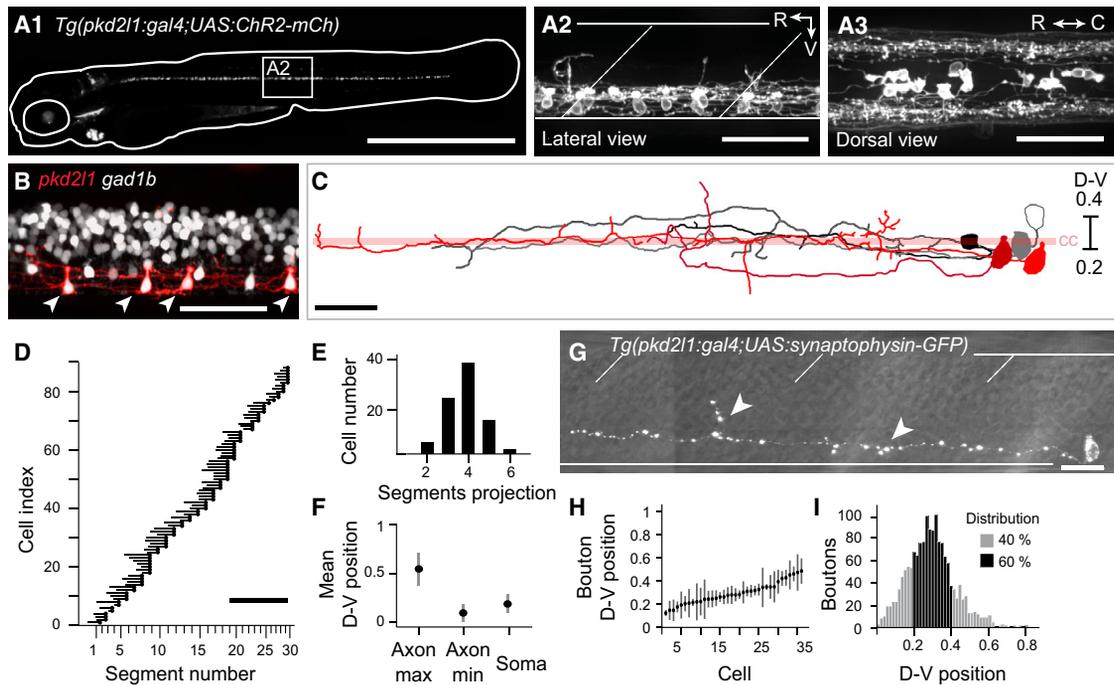


Figure 1. The *Tg(pkcd211:gal4)^{icm10}* Stable Transgenic Line Specifically Labels CSF-cNs in the Zebrafish Spinal Cord

(A1) Complete pattern of expression of mCherry in *Tg(pkcd211:gal4;UAS:ChR2-mCh)* double transgenic larvae at 4 dpf.

(A2) Lateral view of the spinal cord shows that mCherry was restricted to CSF-cNs in the ventral part of the spinal cord. Seven CSF-cNs were labeled per axial segment on average.

(A3) Dorsal view of the spinal cord shows that axonal projections of CSF-cNs were ipsilateral and located in the lateral margins of the spinal cord. See also Figure S1 and Table S1.

(B) Overlap of mCherry (red) and GFP (gray) in the *Tg(pkcd211:gal4;UAS:ChR2-mCh;Gad1b:GFP)* triple transgenic larvae confirms the GABAergic nature of CSF-cNs (arrowheads). Note that the *Tg(pkcd211:gal4)* line does not label all CSF-cNs.

(C) Morphology of five CSF-cNs located in segment 9 at 3 dpf after single-cell imaging and reconstruction. Axonal projections varied in length, in branching as well as in dorsoventral (D-V) positioning. Cells were aligned according to the D-V position of their cell body. The central canal (cc) is represented by the light red bar.

(D) Mapping of CSF-cNs axonal projections across the rostrocaudal (R-C) axis. All CSF-cNs had ascending projections reaching from two to six segments away from the cell body (black circles, $n = 88$ cells).

(E) Distribution of the number of segments covered by single axons of CSF-cNs.

(F) Mean D-V positions of axons (maxima and minima) and soma of CSF-cNs.

(G) Single Synaptophysin-GFP CSF-cN with punctate synaptophysin clustering consistent with presynaptic boutons distributed along the entire axonal arborization (arrowheads). See also Movie S1.

(H) D-V position of putative synaptic boutons for CSF-cNs sorted according to their soma position along the D-V axis ($n = 36$ cells).

(I) Distribution of putative synaptic boutons along the D-V axis (black bars indicate that 60% of the boutons are confined in the 0.2–0.4 interval).

In (A2) and (G), white solid lines delineate the ventral and dorsal limits of the spinal cord; dashed lines indicate the limits of axial segments. Scale bars are 1 mm in (A1), 50 μm in (A2), (A3), and (B), 900 μm in (D), and 20 μm in (C) and (G). R, rostral; C, caudal; V, ventral; CC, central canal. (A)–(B) were reconstructed from Z projection stacks through the entire spinal cord. Data are represented here as mean \pm SD.

locomotor CPG in multiple vertebrate species [15–19] and are selectively active during slow locomotion in zebrafish [17–19]. We further dissected the modulatory role of CSF-cNs onto the slow CPG during specific network states. We confirmed that activating CSF-cNs induced delayed slow locomotor activity at rest. In contrast, the activation of CSF-cNs during ongoing locomotor activity inhibited slow locomotion, reducing the duration and the frequency of locomotor events. Moreover, selective activation of rostral CSF-cNs during ongoing activity led to the silencing of activity along the full length of the spinal cord, indicating that CSF-cNs primarily inhibit descending interneurons. Altogether, our results demonstrate that a single type of conserved GABAergic sensory neuron can tune the excitability of the locomotor CPG in a state-dependent manner, by modulating key excitatory premotor interneurons.

RESULTS

CSF-cNs Are Local, Ipsilateral, and Ascending GABAergic Sensory Neurons that Innervate the Ventrolateral Spinal Cord

CSF-cNs have been recently shown to selectively express the transient receptor potential channel (TRP) polycystic kidney disease 2-like 1 (Pkd211) [20–22]. To analyze the functional morphology underlying the connectivity of CSF-cNs, we cloned the *pkcd211* promoter and generated a specific Gal4 line (Figure 1). Fluorescent in situ hybridization against *pkcd211* revealed that the *Tg(pkcd211:gal4)* line recapitulates the endogenous *pkcd211* expression profile at 3 days post fertilization (dpf) (Figure S1). In *Tg(pkcd211:gal4;UAS:ChR2-mCh)* double transgenic larvae, we obtained selective expression of Channelrhodopsin-2

(ChR2) in CSF-cNs throughout the entire spinal cord (Figure 1A1). CSF-cNs are characterized by an apical villiated extension contacting the central canal and an elongated soma (Figures 1A2 and 1C) [23]. We found that these cells had rostrally directed projections chiefly restricted to the ventrolateral spinal cord (Figures 1A2, 1A3, 1C, 1D, and 1F). We confirmed the GABAergic nature of *pkd211*⁺ CSF-cNs [12, 13, 21, 24] by quantifying the overlap between glutamic acid decarboxylase (Gad) expression and Pkd211 in *Tg(pkd211:gal4;UAS:ChR2-mCh;gad1b:GFP)* larvae (Figure 1B; 99% of mCherry⁺ CSF-cNs were GFP⁺, n = 504 cells in 5 larvae).

We used sparse genetic labeling (see Supplemental Experimental Procedures) to determine the precise projection patterns of CSF-cNs axons. At 3 dpf, all CSF-cNs axons were ascending, ipsilateral, and produced local projections reaching from two to six segments away from the cell body (Figures 1C–1E; n = 88 cells in 62 larvae, mean segment projection length = 3.8 ± 0.8 segments). Regarding the extent of projections within the dorsoventral (D-V) axis, we observed that the axons of CSF-cNs mainly ran in the ventral spinal cord (Figures 1C and 1F; mean D-V axon position = 0.32 ± 0.09 where the ventral limit is 0 and the dorsal limit is 1). To map synaptic sites along the CSF-cN axons, we drove expression of Synaptophysin-GFP to visualize putative presynaptic boutons [25]. Boutons were identified as large and stable GFP⁺ clusters, while small Synaptophysin-GFP-containing vesicles were dim and highly mobile (Figure 1G; Movie S1). The majority of putative synaptic sites was confined in a ventral domain between 0.2 and 0.4 on the D-V axis (Figure 1I; 60% of boutons, see the black portion of the distribution), although presynaptic boutons were observed throughout the entire CSF-cNs D-V axonal projection domain (0–0.6 on the D-V axis; Figures 1H and 1I; n = 1,566 boutons from 36 cells in 27 larvae, mean bouton D-V position = 0.293 ± 0.128). This observation suggests that the primary targets of CSF-cN axons lie within the ventral spinal cord.

CSF-cNs Form GABAergic Synapses onto Premotor V0-v Glutamatergic Interneurons

The cell bodies of a subset of glutamatergic V0-v interneurons referred to as multipolar commissural descending interneurons (MCoDs) in larval zebrafish precisely reside in the ventrolateral spinal cord, confined between 0.2 and 0.4 on the D-V axis [17, 19, 26, 27]. To test whether CSF-cNs project onto glutamatergic V0-v interneurons, we took advantage of the *Tg(vglut2a:lox:DsRed:lox:GFP)* transgenic line labeling most of the glutamatergic neurons in the spinal cord [28, 29]. In this line, we identified the large soma of DsRed⁺ ventrolateral excitatory interneurons at the interface of the lateral neuropil and the cluster of spinal interneurons (Figures 2A1 and 2A2). In *Tg(pkd211:gal4;UAS:ChR2-YFP; vglut2a:lox:DsRed:lox:GFP)* larvae, axonal projections of CSF-cNs formed numerous boutons apposed onto the somata of *vglut2a*⁺ ventrolateral interneurons (top and bottom panels in Figures 2A1, 2A2, and 2B), suggesting that CSF-cNs synapse onto these cells. Moreover, we observed that single CSF-cN axons could project onto multiple ventrolateral *vglut2a*⁺ interneurons (Figure 2B, arrowheads).

We performed targeted whole-cell recordings to measure the electrophysiological properties of ventrolateral *vglut2a*⁺ interneurons with Alexa 647 dye in the recording pipette to reconstruct their morphology (Figure 2C). The morphology and firing

patterns of excitatory ventrolateral *vglut2a*⁺ interneurons were similar to those described previously for glutamatergic V0-v interneurons known as MCoDs in zebrafish (Figures 2D–2G) [17, 26, 27, 30, 31]. Post hoc reconstructions of dye-filled cells demonstrated two dendrites symmetrically located on each side of the soma (Figures 2D and 2E; n = 12 cells) and multiple dendritic ramifications (Figures 2D and 2E). Analysis of presynaptic boutons made by *pkd211*-expressing cells onto dye-filled V0-v interneurons in *Tg(pkd211:Gal4;UAS:ChR2-YFP)* larvae revealed dual innervation of soma and dendrites by the axons of CSF-cNs (Figure 2D, arrowheads).

We made simultaneous whole-cell current clamp recordings of *vglut2a*⁺ V0-v interneurons while monitoring pooled motor output from ventral nerve root (VNR) recordings made from nearby body segments. These dual recordings showed that V0-v cells are rhythmically active during episodes of spontaneous fictive slow locomotion (15–30 Hz, Figure 2F, n = 5). In zebrafish, interneurons active during slow swimming typically exhibit input resistance greater than 400 M Ω [19]. We found that *vglut2a*⁺ V0-v cells exhibited high input resistance (641 ± 52 M Ω , n = 8 cells) and that action potentials could be elicited with small depolarizing currents (Figure 2G; mean threshold current = 15.4 ± 8.3 pA, n = 12 cells), suggesting that these cells are highly excitable and recruited with relatively small levels of excitatory drive [17, 19].

To test whether CSF-cNs formed functional monosynaptic connections onto *vglut2a*⁺ V0-v interneurons, we elicited single spikes in ChR2-expressing CSF-cNs using brief optical activation while recording inhibitory currents in nearby *vglut2a*⁺ V0-v interneurons (Figure 3A1). Pulses of 5 ms blue (~460 nm) light reliably elicited single spikes in CSF-cNs (Figure 3A2; spike delay = 4.86 ± 0.50 ms, n = 141 stimulations in 4 cells). Following single light pulses, we observed GABAergic-mediated inhibitory postsynaptic currents (IPSCs) in V0-v interneurons after a short delay (Figures 3B, 3B1, and 3C; IPSC delay = 4.94 ± 2.02 ms, time to peak = 1.58 ± 0.65 ms, n = 8 cells) and a decay time consistent with the deactivation of GABA_A receptors (IPSC time decay = 25 ± 10.2 ms; [32]). Intriguingly, we found that the probability of observing an IPSC in response to each light pulse was initially low (Figure 3C; response probability = 0.18 ± 0.04 , n = 8 cells) but increased during 500-ms train stimulations at 25 Hz to reach up to 0.5 (Figures 3B2 and 3E; n = 5 cells). The probability of eliciting light-evoked IPSCs increased both during a train (Figures 3D and 3E; p < 0.001, n = 5 cells) and across trains over the time course of the experiment (Figures 3D and 3F; p < 0.001, n = 5 cells), suggesting that individual CSF-cN presynaptic terminals may have a low release probability that is overcome with repeated activation. ChR2-induced IPSCs were blocked upon bath application of gabazine (Figure 3G; n = 3 cells), indicating that CSF-cNs form GABA_A-mediated synapses onto V0-v interneurons. Together, these results indicate that CSF-cNs form active GABAergic synapses onto glutamatergic descending V0-v interneurons that are gradually recruited with repetitive stimulations.

CSF-cNs Exert a State-Dependent Modulation of the Slow Locomotor CPG

Knowing that CSF-cNs project onto premotor excitatory interneurons specifically active during slow locomotion, we tested

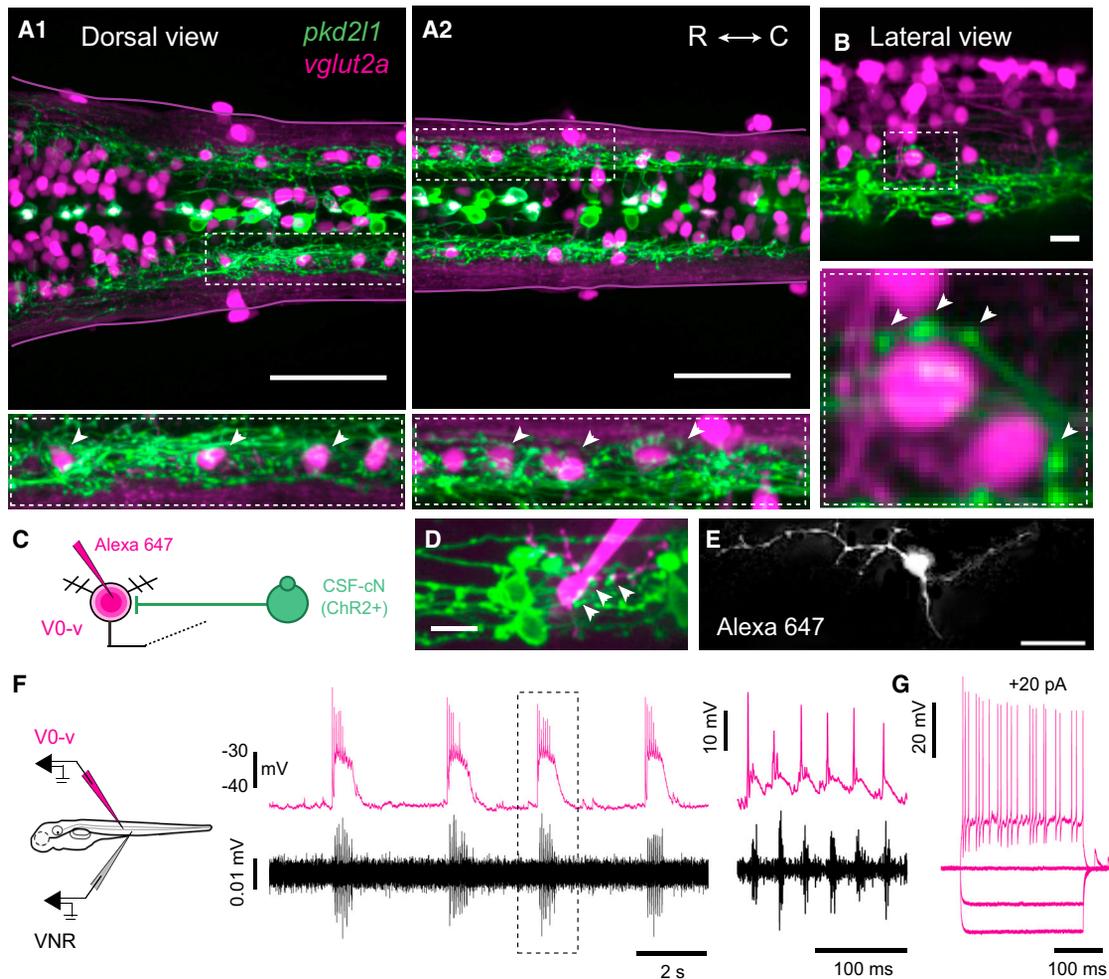


Figure 2. CSF-cNs Project onto V0-v Glutamatergic Interneurons

(A1 and A2) Z projection stack of a few optical sections imaged from the dorsal side in *Tg(pkcd211:gal4;UAS:ChR2-YFP; vglut2a:lox-DsRed-lox-GFP)* triple transgenic larvae reveal the localization of glutamatergic neurons (DsRed⁺, magenta) relative to CSF-cN projections (YFP⁺, green) in rostral (A1) and caudal (A2) spinal cord. Zoom of lateral regions circled in dashed lines show that CSF-cN projections surround the DsRed⁺ nucleus of ventrolateral glutamatergic interneurons (arrowheads in bottom panels, A1 and A2). See also [Table S1](#).

(B) Apposition of CSF-cN axonal varicosities onto the cell bodies of two ventrolateral *vglut2a*⁺ interneurons (arrowheads).

(C) Ventrolateral *vglut2a*⁺ V0-v interneuron receiving projections from CSF-cNs were filled with Alexa 647 in order to image and reconstruct their morphology.

(D) Z projection stack of a few optical sections imaged from the lateral side in a *Tg(pkcd211:gal4;UAS:ChR2-YFP;vglut2a:lox-DsRed-lox-GFP)* triple transgenic larva labeling CSF-cNs (ChR2-YFP⁺, green) and glutamatergic interneurons (DsRed⁺, magenta) after dye filling a *vglut2a*⁺ V0-v interneuron. Arrowheads highlight axonal projections of CSF-cNs onto the soma and dendrites of the filled V0-v interneuron.

(E) Typical morphology of ventrolateral *vglut2a*⁺ V0-v interneurons filled with Alexa 647.

(F) Paired ventral nerve root recording (VNR) with whole-cell current clamp recording of a V0-v interneuron showing rhythmic activity during every episode of fictive slow locomotion (burst frequency ranged between 15 and 30 Hz). The region circled with dashed line is zoomed in on the right to emphasize the activity of V0-v during a fictive bout. V0-v action potentials preceded motor neurons spiking for all locomotor bursts.

(G) V0-v cells are electronically compact, typically requiring only 10–20 pA of current injection to reach action potential (AP)-firing threshold. Note that the cell depicted in (G) is not the same cell as in (F).

Scale bars are 50 μ m in (A1) and (A2) and 10 μ m in (B), (D), and (E). In each panel, Z projection stacks were reconstructed from a few optical sections (depth = 0.55 μ m).

the effects of activating CSF-cNs on locomotor activity. We carefully restrained illumination to the spinal cord (see [Experimental Procedures](#)) and used a long light pulse (500 ms; [Figures 4A and 4B](#)) to elicit burst spiking in ChR2-expressing CSF-cNs (response delay = 7.9 ± 5.8 ms, 11.9 ± 4.3 spikes per light pulse, firing frequency = 23.8 ± 8.7 Hz, burst duration = 491.5 ± 10.6 ms, $n = 2$ cells). In this configuration, blue light stimulation

did not trigger locomotor activity in ChR2⁻ control siblings ([Figure 4C](#), top trace; [Figure 4D](#); response rate = $3.6\% \pm 1.6\%$, $n = 586$ stimulations in 10 larvae). In contrast, there was a significantly higher response rate following the light pulse in ChR2⁺ larvae ([Figure 4C](#), middle trace, and [Figure 4D](#); response rate = 19.96% , $n = 759$ stimulations in 14 larvae, $p < 0.05$). Among the 14 ChR2⁺ larvae tested, we observed that blue light pulses

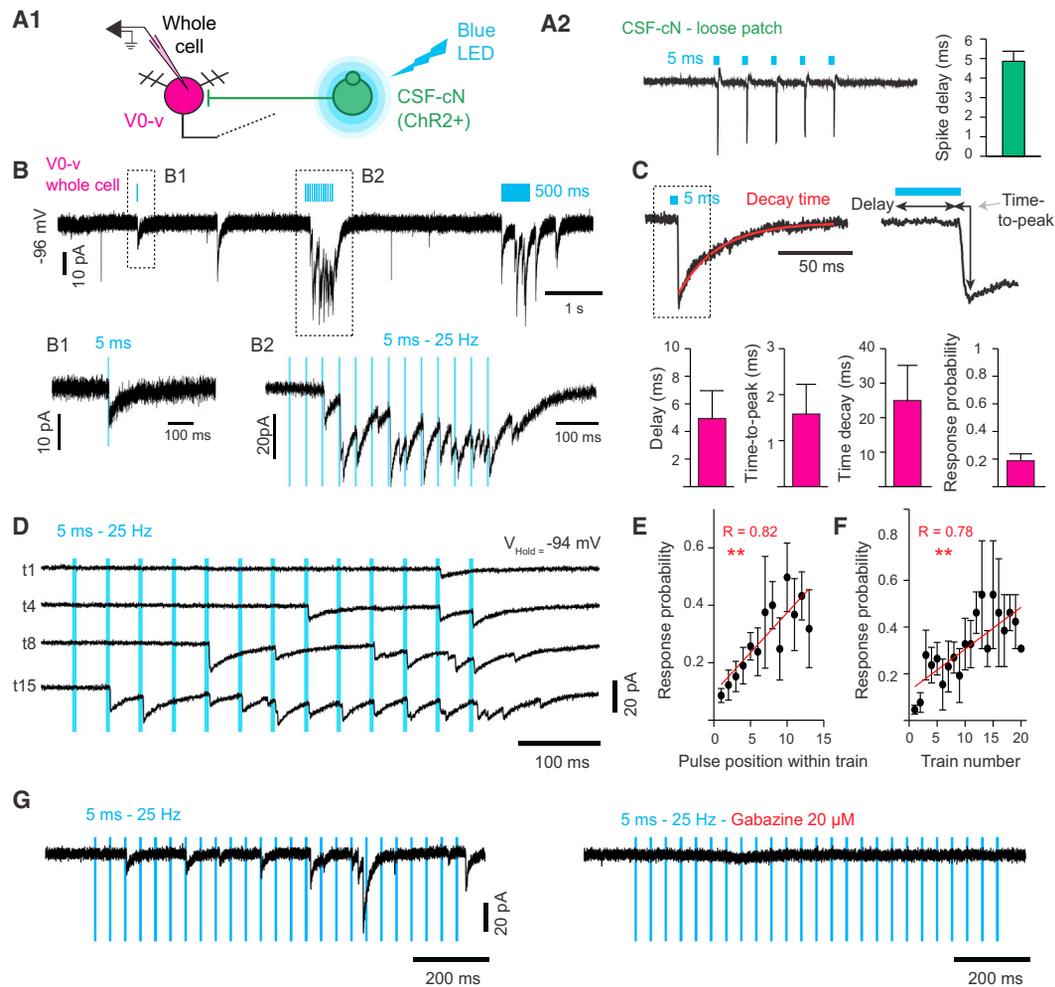


Figure 3. CSF-cNs Form Active GABAergic Synapses onto V0-v Glutamatergic Interneurons

- (A1) Experimental paradigm. Targeted whole-cell voltage-clamp recordings from *vglut2a*⁺ V0-v interneurons while activating ChR2⁺ CSF-cNs using brief (5 ms) pulses of blue light.
- (A2) Loose-patch recording of a ChR2⁺ CSF-cN shows that 5-ms blue light pulses (blue bars) reliably triggered single action potentials in CSF-cNs. The plot indicates that the spike delay relative to the onset of the light pulse was about 4–5 ms (141 stimulations from 4 cells).
- (B) Voltage clamp recording of a V0-v interneuron during optical stimulation of ChR2⁺ CSF-cNs (blue bars) using a single pulse (B1) and a train of 13 pulses at 25 Hz (B2). Holding potential $V_m = -96$ mV and reversal potential for chloride $E_{Cl} = -51$ mV; therefore, IPSCs appear as inward currents.
- (C) Quantification of kinetic parameters of ChR2-induced IPSCs in V0-v interneurons; delay was calculated relative to the onset of the light pulse, decay time was obtained after fitting the decay with a single exponential fit, and time to peak was calculated as the difference between the time of current onset and of current peak.
- (D) Reliability of IPSC events increased for subsequent pulses during a trial as well as across subsequent trials recorded every 20–25 s (1st, 4th, 8th, and 15th trials shown).
- (E) Quantification of the response probability in V0-v interneurons for each pulse during a train.
- (F) Quantification of the response probability in V0-v interneurons, averaged per pulse during each train, as a function of the train number over the time course of the experiment.
- (G) ChR2-induced IPSCs in V0-v interneurons are blocked by application of gabazine in the bath ($n = 3$ cells). Blue bars represent the stimulation at 25 Hz. Red lines in (E) and (F) are linear fits. ** $p < 0.001$ following Pearson's linear correlation test. R is the linear correlation coefficient. Data are represented as mean \pm SEM.

did not induce locomotor activity in six animals (Figure 4C, bottom trace; response rate = $1.8\% \pm 0.8\%$). In 8 out of 14 ChR2⁺ larvae, the activation of CSF-cNs reliably triggered slow fictive swimming after a delay of 465 ± 55 ms (Figure 4C, middle trace; response rate = $33.6\% \pm 8.4\%$). Such a long delay suggested that locomotor responses followed an initial period of inhibition. We tested whether the induced locomotor response was GABA

mediated on a larva with a high baseline response rate. Bath application of the GABA_A receptor antagonist gabazine led to a reduction of the response rate from 0.79 to 0.16, suggesting that the rebound swimming could rely on the activation of GABA_A receptors in this animal (Figure 4E).

We hypothesized that the heterogeneity of the responses observed across ChR2⁺ larvae could be due to variations in

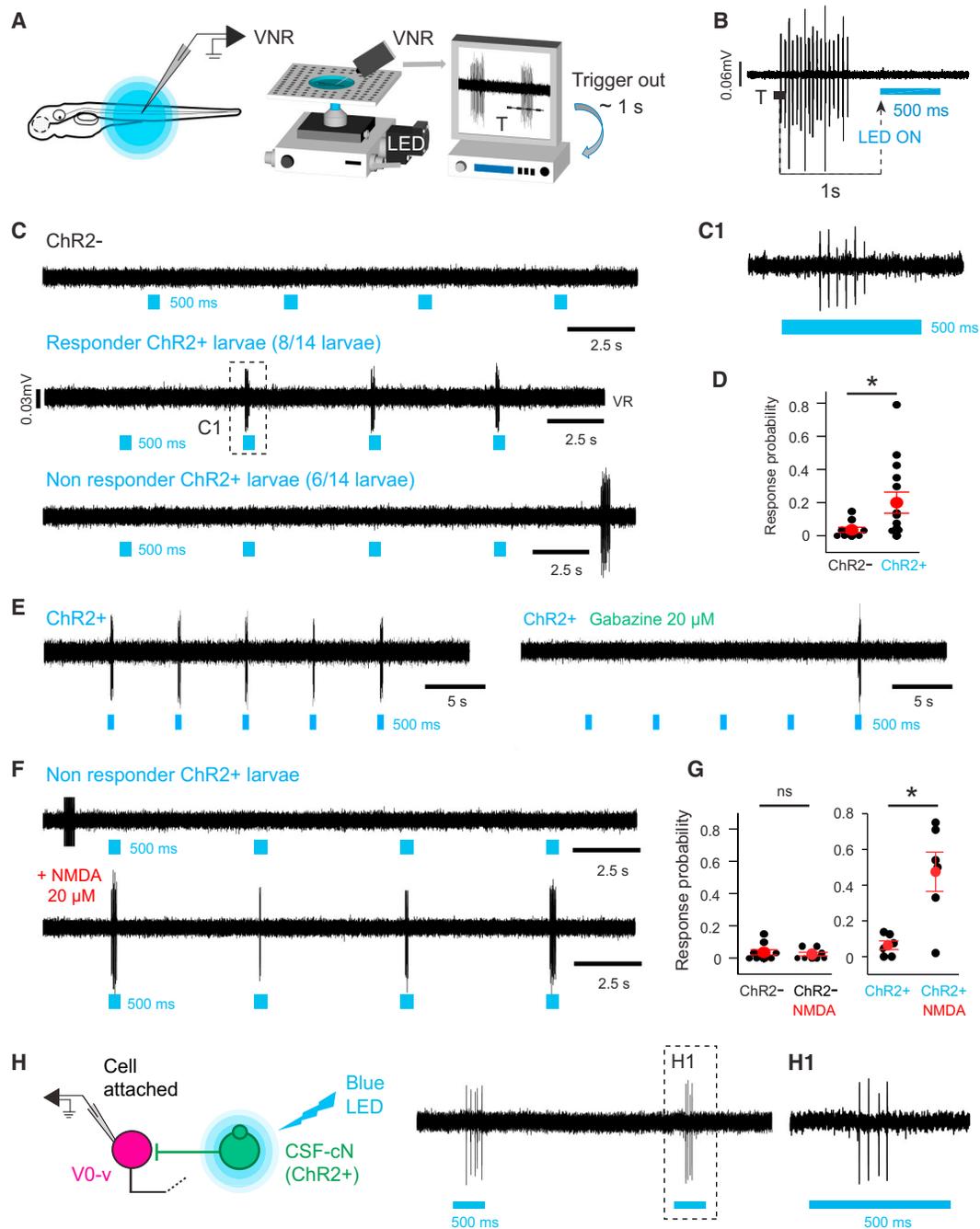


Figure 4. Activation of CSF-cNs at Rest Can Trigger Delayed Slow Locomotion

(A) Experimental paradigm showing an eye-enucleated 4 dpf ChR2⁺ larva mounted on its side in a glass-bottom dish after paralysis. Blue light was patterned onto the spinal cord from segment 7–8 to 27–28 through the microscope condenser. VNR signals were recorded from the axial musculature and analyzed in real time. A threshold (T) was set to trigger the LED based on the VNR signal.

(B) Photoactivation protocol. LED was triggered during resting period when the animal was not fictively swimming. If the larva was not swimming, the LED was automatically activated every 4–5 s. If the larva was swimming, the LED was triggered long (500 ms to 2 s) after the onset of fictive bouts. The blue bar represents the pulse of blue light.

(C and C1) Sample traces of VNR recordings showing the effects of activating ChR2⁺ CSF-cNs using blue light at rest (light pulses are represented by the blue bars). ChR2⁻ control siblings never showed swimming activity in response to blue light stimulations (top, n = 10 larvae). A responding ChR2⁺ responder larva showed a delayed swimming response after the onset of the light pulse (middle, n = 8 out of 14; zoom in C1). Some ChR2⁺ larvae did not respond to the light stimulation at rest (bottom, n = 6 out of 14).

(D) Quantification of the probability to induce delayed fictive swimming after blue light stimulations in ChR2⁻ and ChR2⁺ larvae (n = 10 and 14, respectively).

(E) Sample VNR traces illustrating that the induction of delayed swimming in a responsive ChR2⁺ larva was blocked after the addition of 20 μM gabazine in the bath.

(legend continued on next page)

the intrinsic excitability of the spinal locomotor circuit across fish. The excitability of spinal circuits can be modulated with the application of excitatory neurotransmitters such as NMDA [33, 34]. Bath application of low concentrations of NMDA (10–20 μ M) had no effect on swimming in ChR2⁻ control siblings (Figure 4G, left plot; response rate = 3.6% \pm 1.6% at rest and 2.4% \pm 1.1% with NMDA, n = 8 larvae). In contrast, the presence of NMDA led to a dramatic increase in the response rate to photostimulation in ChR2⁺ larvae, by converting larvae with no response into responsive larvae (Figure 4F, compare top and bottom traces; Figure 4G, right plot; the response rate went from 6.4% \pm 2.4% at rest to 47% \pm 11% under NMDA, n = 6 larvae, p < 0.05).

We tested whether *vglut2a*⁺ V0-v interneurons contributed to the delayed motor activity following the activation of CSF-cNs at rest. We recorded V0-v in cell-attached mode and analyzed their firing in response to blue light (Figure 4H). We found that activating CSF-cNs could induce firing in V0-v (response delay = 320 \pm 167 ms, n = 118 out of 311 stimulations in 3 cells). Altogether, these data indicate that the induction of slow locomotion at rest following CSF-cN activation involves the recruitment of V0-v interneurons.

Because CSF-cNs form active GABAergic synapses onto glutamatergic V0-v premotor interneurons, we hypothesized that CSF-cN activation during ongoing locomotion might silence motor activity. We used a closed-loop ChR2 activation assay (Figures 5A and 5B) where a 500-ms light pulse was delivered at the onset of spontaneous slow fictive swimming events. In ChR2⁻ control siblings, blue light pulses had no effect on ongoing locomotor activity (Figure 5C). In contrast, activation of ChR2⁺ CSF-cNs led to an abrupt silencing of ongoing swimming activity (Figure 5D). The activation of CSF-cNs significantly reduced the duration of spontaneous swim bouts (Figures 5E–5G; for control fish, bout duration with light-emitting diode (LED) off [BD_{LED OFF}] = 550 \pm 53 ms, BD_{LED ON} = 486 \pm 59 ms, n = 15 larvae; whereas for ChR2-expressing fish, BD_{LED OFF} = 840 \pm 82 ms versus BD_{LED ON} = 235 \pm 33 ms, p < 0.001, n = 29 larvae). In addition, CSF-cN activation led to a significant reduction in the frequency of swim bouts (Figure 5H; for control fish, bout frequency with LED off [BF_{LED OFF}] = 0.5, BF_{LED ON} = 0.56, n = 7 larvae; for ChR2-expressing fish, BF_{LED OFF} = 0.52, BF_{LED ON} = 0.33, p < 0.05, n = 6 larvae) without altering the burst frequency (bf) within bouts (Figure S2; bf_{LED OFF} = 22.60 \pm 0.64 Hz, bf_{LED ON} = 22.54 \pm 0.80 Hz). These findings demonstrate a marked effect of CSF-cN activation on the duration and occurrence of spontaneous bouts of fictive swimming.

We tested whether the silencing of locomotor activity was mediated by the activation of ionotropic GABA_A receptors. Bath application of GABA_A blocker gabazine (10–20 μ M) led to an increase in the spontaneous bout duration (compare the left panel of Figure 5I with the left panel of Figure 5J; Figure 5K,

BD_{LED OFF} = 909 \pm 117 ms, BD_{LED OFF/gabazine} = 2.18 \pm 0.64 s, p < 0.0001, n = 10 larvae), indicating that the endogenous release of GABA modulates the duration of locomotor events. Nonetheless, the presence of gabazine reduced the silencing mediated by CSF-cNs (compare Figure 5I right panel with Figure 5J right panel; Figure 5K, BD_{LED ON} = 184 \pm 38 ms, BD_{LED ON/gabazine} = 779 \pm 128 ms, p < 0.0001). Since the silencing of CSF-cNs was independent of the initial bout duration (Figure S3), we measured the silencing efficiency of CSF-cNs, i.e., their ability to reduce bout duration, as the ratio of bout duration with or without the activation of ChR2⁺ CSF-cNs (BD_{LED OFF}/BD_{LED ON}). The silencing efficiency was reduced from 6.9 to 2.9 when gabazine was added into the bath (Figure 5L; p < 0.05, n = 10), indicating that the inhibition of fictive locomotion involves the activation of GABA_A receptors.

Together, these results demonstrate that CSF-cNs can modulate the slow locomotor CPG in a state-dependent manner. CSF-cNs can trigger delayed rhythmic activity at rest, an effect enhanced by 10–20 μ M NMDA in the bath. In contrast, CSF-cNs can inhibit ongoing locomotor activity, reducing both the duration and the frequency of occurrence of locomotor events.

Inhibition of Locomotor Activity by CSF-cNs Predominantly Occurs at the Premotor Level

The suppression of ongoing locomotor activity could be due to the direct inhibition of motor neurons and/or of descending premotor excitatory interneurons controlling slow locomotion. We first tested whether CSF-cN activation could silence ongoing activity in V0-v excitatory interneurons recorded in cell-attached mode (Figure 6A). We observed that spontaneous firing events in V0-v interneurons were silenced by CSF-cN activation (Figure 6B) leading to shorter episodes (Figures 6C and 6D; mean event duration with LED off [ED_{LED OFF}] = 341 \pm 50 ms, ED_{LED ON} = 176 \pm 77 ms, n = 99 and 77 episodes, respectively, recorded in n = 3 cells) containing fewer spikes (Figure 6E; mean spikes per episode with LED_{OFF} = 15.8 \pm 4.9 spikes, mean spikes per episode with LED_{ON} = 7.7 \pm 2.8 spikes). These data reveal that CSF-cNs can silence locomotor activity at the premotor level.

Since glutamatergic V0-v interneurons project 15–20 segments caudally [17, 27] and are silenced by CSF-cNs, we hypothesized that silencing rostral V0-v interneurons could alter the propagation of locomotor activity to more caudal motor pools. To test this hypothesis, we restricted the optical activation of CSF-cNs to rostral (1–10) or caudal (16–25) segments while performing dual VNR recordings at rostral (5–8) and caudal (15–16) segments (Figures 7A–7C). The activation of rostral CSF-cNs strongly silenced fictive swimming activity in rostral as well as in caudal segments, occasionally abolishing caudal motor activity (Figure 7B, see stars on bottom traces; Figure 7D; n = 6 larvae). In contrast, the activation of caudal

(F) Sample traces showing that 20 μ M NMDA bath application transformed ChR2⁺ non-responder larvae into responder larvae. Note that after NMDA application, the light response of the ChR2⁺ non-responder larvae mimicked the response observed in responder larvae (C, middle).

(G) Quantification of the probability to induce delayed fictive swimming responses in ChR2⁻ and ChR2⁺ larvae before and after NMDA application.

(H and H1) Targeted cell attached recordings from a *vglut2a*⁺ V0-v interneuron while activating CSF-cNs expressing ChR2 using 500-ms pulses of blue light. Delayed firing is triggered by blue light stimulations. 118 out of 311 stimulations led to delayed firing in n = 3 V0-v cells. The circled region is zoomed in on (H1) to better resolve the spiking.

*p < 0.05 following Student's t test. Data are represented as mean \pm SEM. Mean values are depicted in red.

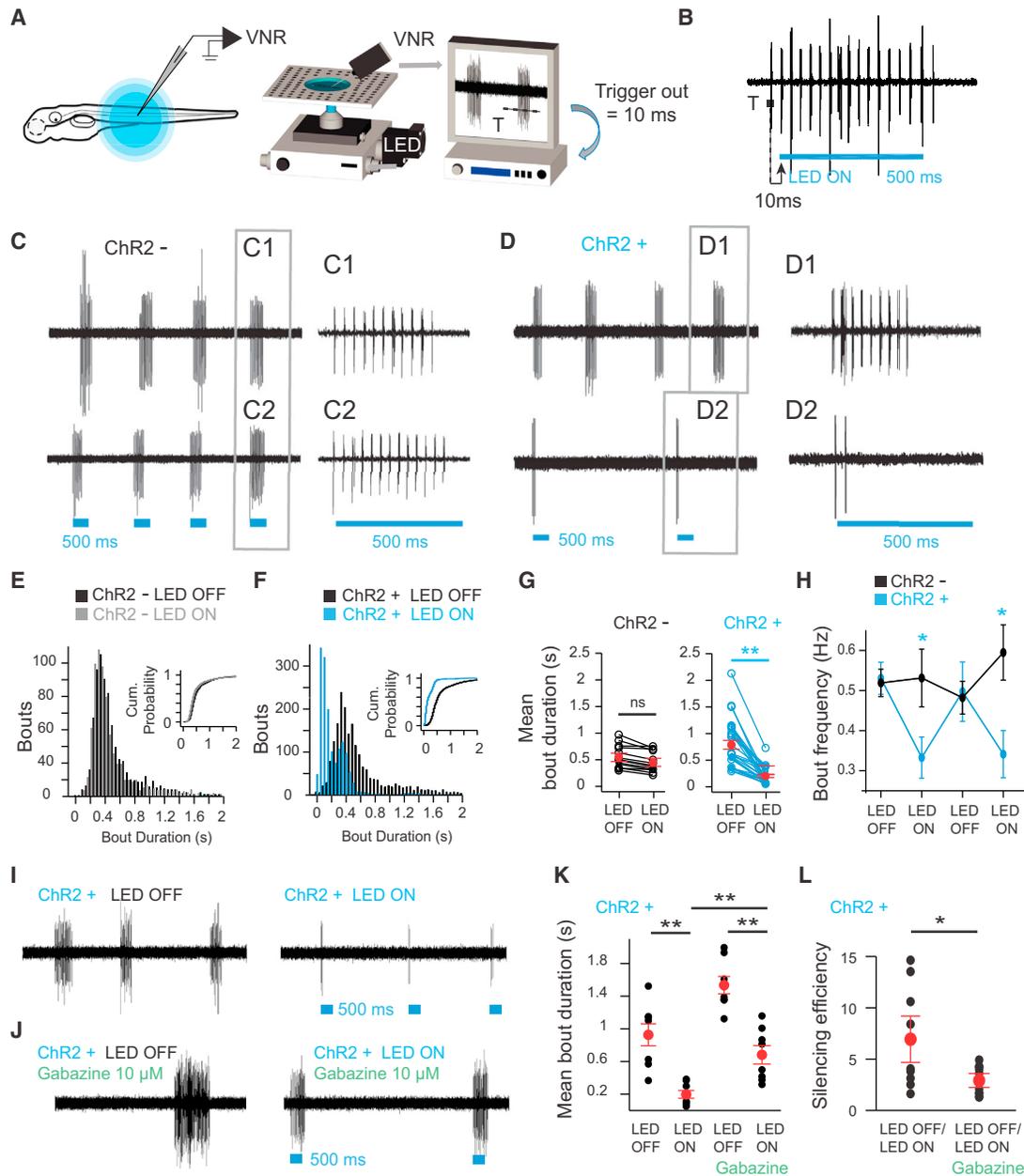


Figure 5. Activation of CSF-cNs at the Onset of Ongoing Fictive Swimming Silences Locomotor Activity

(A) Experimental paradigm; same as in Figure 4A except that the LED was triggered at the onset of each fictive swim.

(B) Photoactivation protocol. LED was triggered rapidly (10 ms) after the onset of spontaneous fictive bouts that were detected when the VNR signal had reached a manually defined threshold (T). The blue bar represents the light pulse.

(C–C2) In ChR2⁻ control siblings, profiles of fictive locomotor activity without (top) or with (bottom) blue light stimulations (blue bars) were similar (zooms in C1 and C2).

(D–D2) Same as in (C) in ChR2⁺ larvae; a reduction of bout duration (BD) and bout frequency (BF) is associated with the optical activation of CSF-cNs (zoom on bouts in D1 and D2).

(E) Distribution of spontaneous fictive bout durations in ChR2⁻ control siblings was similar with LED_{OFF} (black bars) or LED_{ON} (gray bars). The inset shows the cumulative probability of fictive bout duration in LED_{OFF} and LED_{ON} conditions.

(F) Same as in (E) in ChR2⁺ larvae. In contrast to ChR2⁻ control siblings, the distribution and cumulative probability were different in LED_{OFF} (black bars) and LED_{ON} (blue bars) conditions, indicating a large decrease of bout duration when the LED was ON.

(G) Quantification of the mean bout duration in both conditions for ChR2⁻ and ChR2⁺ larvae.

(H) Quantification of the bout frequency (BF) in ChR2⁻ and ChR2⁺ larvae in recordings where the LED was alternatively ON and OFF. In control ChR2⁻ larvae, the frequency did not change over the time course of the experiment, whereas the bout frequency reversibly decreased with LED_{ON} in ChR2⁺ larvae. See also Figure S2.

(I) Sample VNR traces in a ChR2⁺ larva illustrating the typical silencing mediated by CSF-cNs.

(legend continued on next page)

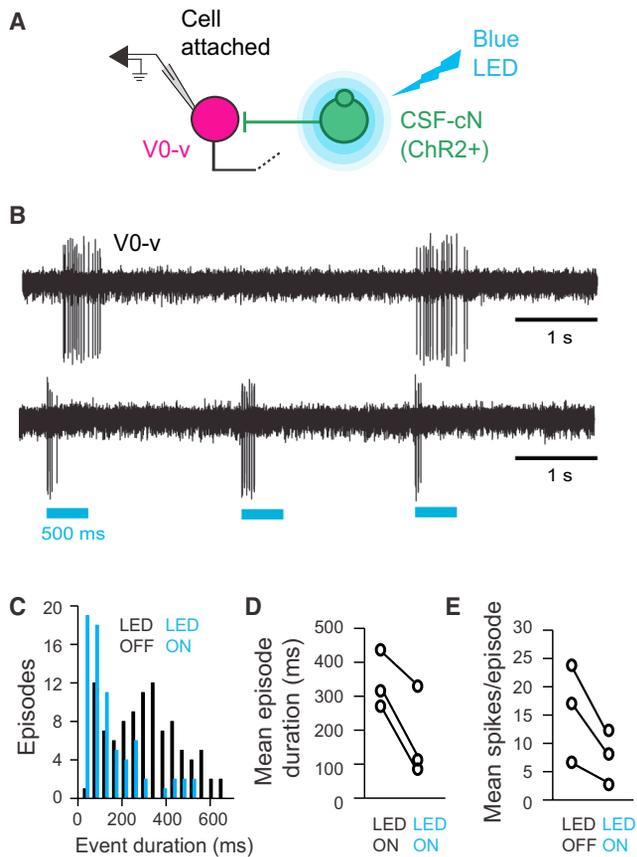


Figure 6. Activation of CSF-cNs Silences Episodes of Glutamatergic V0-v Interneuron Activity

(A) Experimental paradigm: targeted cell attached recordings from *vglut2a*⁺ V0-v interneurons while activating CSF-cNs expressing ChR2 using 500-ms pulses of blue light.

(B) Episodes of V0-v activity (top: spontaneous activity; bottom: with stimulation of CSF-cNs timed to the onset of V0-v episodes). Blue bars represent blue light pulses.

(C) Distribution of V0-v episode durations with LED_{OFF} or LED_{ON}.

(D) Mean episode duration per cell with LED_{OFF} or LED_{ON}.

(E) Mean number of spikes per episode per cell with LED_{OFF} or LED_{ON}.

CSF-cNs had small effects on both rostral and caudal motor activity compared to the activation of rostral CSF-cNs (Figures 7C and 7D; $n = 4$ larvae; Figure 7E; silencing efficiency for rostral stimulations = 2.85 at VR 8 and 2.04 at VR 16, silencing efficiency for caudal stimulations = 1.08 at VR 8 and 1.06 at VR 16, $p < 0.01$). The activation of rostral CSF-cNs did not seem to modulate the rostrocaudal lag measured between segment 8 and 16 (Figure S4). Altogether, these results suggest that the ability of CSF-cNs to disrupt the excitatory drive along the spinal cord most likely relies on the modulation of premotor in-

terneurons as these cells have long descending projections within the spinal cord.

DISCUSSION

In the present study, we investigated the cellular and network mechanisms underlying the modulation of slow locomotion by CSF-cNs in larval zebrafish. To our knowledge, this work is the first investigation of the functional connectivity of a single GABAergic neuron type onto glutamatergic interneurons of the locomotor CPG. Our results highlight the complexity of this GABAergic modulatory pathway leading to antagonistic effects depending on the excitability or state of spinal motor circuits.

Circuit Organization of Spinal CSF-cNs: Projections onto Excitatory Elements of the Slow Swimming CPG

Taking advantage of existing transgenic lines labeling glutamatergic interneurons [28, 29], we observed anatomical and functional connections from CSF-cNs onto *vglut2a*⁺ V0-v premotor interneurons in the spinal cord. In larval zebrafish, these cells are specifically active during slow locomotion with bursting frequencies ranging from 15 to 30 Hz and are selectively silenced at swimming frequencies above 30 Hz, possibly by glycinergic interneurons [17]. Our results demonstrate that additional GABAergic inputs to *vglut2a*⁺ V0-v originate from CSF-cNs. Given the increasing fidelity of ChR2-driven IPSCs between CSF-cNs and V0-v during 25-Hz train stimulations, CSF-cNs may also contribute to the frequency-dependent suppression of V0-v activity. In mice, although glutamatergic V0-v interneurons are essential components of the locomotor CPG [15, 16], they appear critical for left-right alternation during fast locomotion [35], and their ablation selectively affects trot [36]. Further work will be necessary in zebrafish to test whether CSF-cNs can modulate left-right alternation.

Interestingly, activation of CSF-cNs reduced the occurrence of locomotor events. This effect could be explained by the modulation of supraspinal neurons thought to control the initiation and frequency of locomotor events [37, 38]. In the rostral spinal cord, we observed that CSF-cNs projected into the lateral margins of the caudal hindbrain, reaching the somata of V0-v interneurons as well as descending excitatory fibers projecting in the spinal cord. CSF-cNs projecting onto the caudal hindbrain could delay the occurrence of locomotor events by silencing the output of hindbrain interneurons projecting in the spinal cord. However, the full connectivity map of CSF-cNs on their targets, including neurons located in the hindbrain, remains to be completed.

Regarding the neurotransmitter released by CSF-cNs, the modulation of locomotion by CSF-cNs was at least partially mediated by GABA_A receptors. However, there might be additional components to the effect mediated by CSF-cNs.

(J) Same as in (I) after bath application of 10 μ M gabazine. Note that bout duration increased upon application. The drug penetrated the spinal cord about 5 min after application. Data were collected between 5 and 15 min after application, before seizures occurred.

(K) Quantification of fictive bout duration in ChR2⁺ larvae during either LED_{OFF} or LED_{ON} episodes as well as before and after gabazine was bath applied.

(L) CSF-cN-mediated silencing efficiency, calculated as the relative reduction of bout duration after CSF-cN activation ($BD_{LED\ OFF}/BD_{LED\ ON}$ ratio), decreased upon gabazine treatment. See also Figure S3.

In (G), (H), and (L), $*p < 0.05$ and $**p < 0.0001$ following Student's *t* test. In (K), a linear mixed-effects model was applied to compare datasets, and post hoc multiple comparisons of means were performed to extract *p* values ($**p < 0.001$). Data are represented as mean \pm SEM. In (G), (K), and (L), mean values are depicted in red.

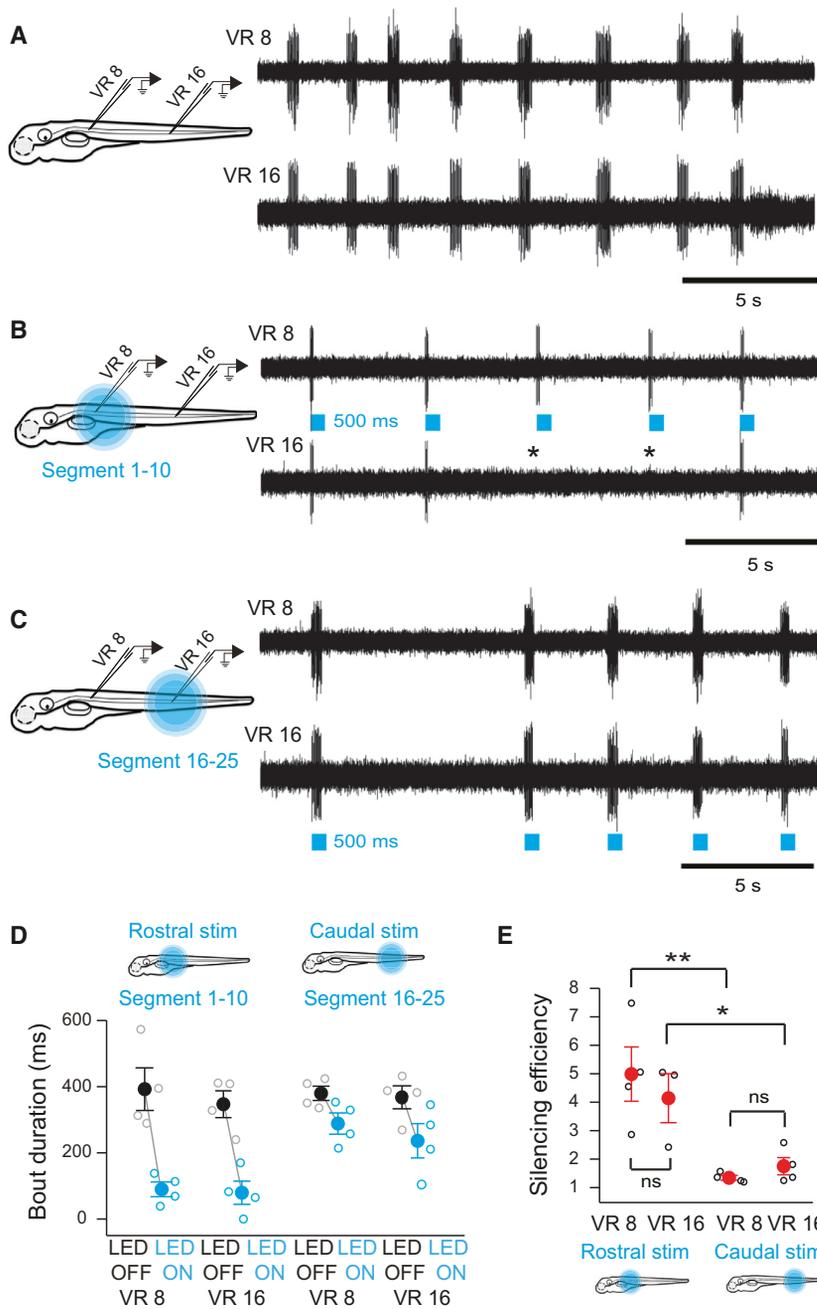


Figure 7. Selective Activation of Rostral CSF-cNs Disrupts the Propagation of Activity to the Caudal Spinal Cord

(A) Profile of spontaneous fictive locomotor activity in Chr2⁺ larvae recorded at rostral (VR8) and caudal (VR16) segments. The field of photoactivation was limited by narrowing the aperture of the microscope condenser.

(B) Fictive locomotor activity was silenced in the rostral VNR as well as in the caudal VNR following the activation of rostral CSF-cNs (segments 1–10). See also Figure S4.

(C) In contrast, fictive locomotor activity in the same larva was not altered following the activation of caudal CSF-cNs (segments 16–25).

(D) Effect of activation of caudal CSF-cNs versus rostral CSF-cNs. Mean bout duration calculated from VNR recordings at segment 8 (VR 8) and segment 16 (VR 16) in LED_{OFF} (black) or LED_{ON} (blue) conditions.

(E) CSF-cN-mediated silencing efficiency, calculated as the relative reduction of bout duration after CSF-cN activation ($BD_{LED\ OFF}/BD_{LED\ ON}$ ratio), measured at segment 8 (VR 8) and 16 (VR16) during rostral or caudal activation of CSF-cNs.

* $p < 0.05$, ** $p < 0.01$ following Student's *t* test. Data are represented as mean \pm SEM.

motor CPGs by CSF-cNs. On one side, we revealed the inhibitory action of CSF-cNs when stimulated during ongoing locomotion. In this context, locomotor activity was silenced within 200 ms on average, suggesting that a buildup of inhibition was necessary. On the other side, activation of CSF-cNs at rest induced delayed fictive swimming that was highly dependent on the intrinsic excitability of the spinal cord. The delay of induced swimming was about 450 ms, ruling out a direct activation of locomotor CPGs. One possible explanation is that rebound activity originates from an accumulation of depolarizing inhibition as depolarizing GABA is common in immature spinal circuits [40]. Alternatively, the induction of swimming may follow a rebound from GABA_A-mediated inhibition. Post-inhibitory rebound (PIR), a general feature of rhythmic networks including locomotor

First, gabazine in the bath failed to fully abolish the silencing of ongoing locomotion. Second, the relatively short inactivation time of GABA_A receptors (<100 ms) does not match the long-lasting effects (seconds) of increasing interbout interval. CSF-cNs have been shown to express a variety of peptides [24, 39]. It is therefore plausible that other receptors for GABA and/or peptides also contribute to the modulation of locomotor activity by CSF-cNs.

State-Dependent Modulation of Locomotor Activity

Using electrophysiology and pharmacology, our study sheds light on the state-dependent GABAergic modulation of loco-

CPGs [41–43], has been proposed to regulate the timing of activation of premotor interneurons [41, 42, 44–46]. In the tadpole, PIR is an emergent property of a complex interplay of inhibition and depolarization that is modulated in a state-dependent manner [43]. Even though our data indicate that V0-v contribute to the delayed activity triggered by CSF-cNs at rest, it is unlikely that PIR originates solely from the intrinsic properties of these interneurons as we did not observe post-hyperpolarization rebound spiking in these cells. Following CSF-cN activation, PIR may rise from network interactions via other targets, leading to the rebound firing of V0-v interneurons and subsequent induction of slow swimming.

Roles for a CSF-Dependent GABAergic Inhibition of Premotor Excitation

Since our experiments relied on forcing the activation of CSF-cNs with light, one open question lies in identifying the physiological conditions and the timing under which these cells are normally recruited *in vivo*. Kolmer initially thought that CSF-cNs could form a parasagittal organ functioning as a third ear in the spinal cord [9]. Indeed, the morphology of these cells extending in the central canal is optimal to detect chemical or mechanical cues from the CSF. Previous studies in mammals indicated that CSF-cN firing was modulated by changes of extracellular pH [20, 47], but how such information is transduced and relates to locomotion is still unclear. The observation that the reliability of CSF-cNs to V0-v synaptic currents increases during 25-Hz train implies that the silencing mediated by CSF-cNs is particularly efficient with persistent spiking in the range of slow swimming frequencies. The inhibition of CSF-cNs onto V0-v interneurons could thereby build up over time during locomotor events. Future work will be necessary to complete the connectivity pattern and modulatory role of CSF-cNs in fish and mammals in order to elucidate CSF-cN modulatory function of active locomotion in vertebrates.

EXPERIMENTAL PROCEDURES

Animal Care

Animal handling and procedures were validated by ICM and the National Ethics Committee (Comité National de Réflexion Ethique sur l'Expérimentation Animale, Ce5/2011/056) in agreement with EU legislation. Adults were reared at a maximal density of eight animals per liter in a 14/10 hr light/dark cycle environment. Fish were fed live artemia twice a day, and feeding regime was supplemented with solid extracts matching the fish developmental stage (ZM Systems). Larvae were raised at 28.5°C with a 14/10 day/night light cycle. Experiments were performed at room temperature (22°C–25°C) on 3–5 dpf larvae.

Generation of Transgenic Animals

Transgenic lines used in this study are listed in Table S1. Detailed procedures including the generation of transgenic animals, FISH combined with IHC, and the morphological analysis of CSF-cNs are available in Supplemental Experimental Procedures.

Live Imaging of Spinal Neurons

Zebrafish larvae were imaged using an upright microscope (Examiner Z1, Zeiss) equipped with a spinning disk head (CSU-X1, Yokogawa) and a modular laser light source (LasterStack, 3i Intelligent Imaging Innovations). Z projection stacks were acquired using Slidebook software (3i) and reconstructed online using Fiji (<http://fiji.sc/Fiji>).

Sample Preparation for Fictive Locomotion Recordings and Optogenetic Stimulations

3 dpf *Tg(pkcd211:gal4);UAS:ChR2-YFP* larvae were screened for dense labeling and bright expression of ChR2-YFP in CSF-cNs under a dissecting fluoroscope (Leica). Larvae were anaesthetized in 0.02% Tricaine-Methiodide (MS-222, Sigma-Aldrich) diluted in fish facility water and then mounted upside-down (ventral side facing up) in glass-bottom dishes (MatTek) filled with 1.5% low-melting-point agarose. We surgically removed the eyes using a thin tungsten pin in order to avoid light-evoked locomotion with blue light during ChR2 stimulation. Following the surgery, larvae were transferred in cold ACSF (concentrations in mM: 134 NaCl, 2.9 KCl, 1.2 MgCl₂, 10 HEPES, 10 glucose, and 2.1 CaCl₂; 290 mOsm, adjusted to pH 7.7–7.8 with NaOH) for 3–5 min. Larvae were then transferred in fish facility water to recover for 24 hr at 28°C. The following day, larvae were mounted on their side and immobilized by injecting 0.5 nl of 0.5 mM α -Bungarotoxin in the ventral axial muscu-

lature (Tocris). A portion of agarose was removed using a sharp razorblade in order to expose 2–3 segments.

Fictive Locomotion Recordings and Optogenetic Stimulation

Our recording protocol is based on published procedures [17, 19, 48]. VNR recordings were acquired using a MultiClamp 700A amplifier, a Digidata series 1322A Digitizer, and pClamp 8.2 software (Axon Instruments, Molecular Devices). A blue LED (UHP-Mic-LED-460, Prizmatix) was used to activate ChR2. The light was delivered on the fish spinal cord through the microscope condenser, typically 20 segments from segment 7–8 to 27–28 with 14 mW/mm². To time the optical activation of ChR2 after the onset of fictive swimming bouts, we designed a closed-loop program in which the LED was turned on via transistor-transistor logic (TTL) pulses 10 ms or 500 ms after the fictive motor output reached an arbitrary threshold. Parameters describing the fictive locomotion were extracted using custom-made MATLAB scripts. Bout frequency was analyzed in larvae with basal level of swimming activity above 0.3 Hz.

Pharmacology

4 dpf larvae were pinned down in Sylgard-coated, glass-bottom dishes filled with ACSF with thin tungsten pins through the notochord. Skin was removed from segments 5–6 to the end of the tail using sharp forceps. GABA_A receptor antagonist gabazine (SR95531, Tocris) or NMDA (Tocris) was bath applied at either 10 or 20 μ M final concentrations.

Fluorescence-Guided Whole-Cell Recordings

Whole-cell recording were performed in head-off larvae in the same configuration as pharmacology experiments. After removing the skin, one to two segments were dissected using glass suction pipettes. Patch pipettes (1B150F-4, WPI) were designed to reach a tip resistance of 11–15 M Ω and were filled with potassium-containing internal solution (concentrations in mM: K-gluconate 115, KCl 15, MgCl₂ 2, Mg-ATP 4, HEPES free acid 10, EGTA 0.5, 290 mOsm, adjusted to pH 7.2 with KOH and supplemented with Alexa 647 at 4 μ M final concentration). To resolve evoked inhibitory postsynaptic currents in voltage-clamp mode, cells were held at around –80 mV, away from the calculated chloride reversal potential (–51 mV). We calculated the liquid junction potential in our experiments (–19 mV) but did not correct for it since it did not affect the outcome of our experiments. Kinetic parameters of light-evoked currents and IPSCs were extracted and analyzed using custom-made MATLAB scripts.

Statistics

Linear correlation in datasets was calculated using a Pearson's linear correlation test. Comparisons between two groups of data were performed using a Student's *t* test. A linear mixed-effects model was used to test the interaction between the LED and gabazine. The level of significance was $p < 0.05$ for all datasets.

ACCESSION NUMBERS

The three new zebrafish transgenic lines *Tg(pkcd211:gal4)^{icm10}*, *Tg(UAS:ChR2-YFP)^{icm11}*, and *Tg(UAS:GCaMP5G)^{icm08}* have been deposited in the Zebrafish Model Organism database under ID codes ZFIN: ZDB-FISH-150901-9831, ZDB-FISH-150901-19823, and ZDB-FISH-150901-6255.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures, four figures, one table, and one movie and can be found with this article online at <http://dx.doi.org/10.1016/j.cub.2015.09.070>.

AUTHOR CONTRIBUTIONS

K.F. performed electrophysiological recordings, pharmacology experiments, and imaging of spinal lines with the help of C.S. L.D. and A.P. performed single-cell morphology analysis. F.D.B. and A.B. generated transgenic animals. J.G. performed FISH experiments. K.F. and C.W. designed experiments, analyzed data, and wrote the manuscript.

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REFERENCES

- Rudomin, P. (1990). Presynaptic inhibition of muscle spindle and tendon organ afferents in the mammalian spinal cord. *Trends Neurosci.* *13*, 499–505.
- Rudomin, P. (2009). In search of lost presynaptic inhibition. *Exp. Brain Res.* *196*, 139–151.
- Betley, J.N., Wright, C.V., Kawaguchi, Y., Erdélyi, F., Szabó, G., Jessell, T.M., and Kaltschmidt, J.A. (2009). Stringent specificity in the construction of a GABAergic presynaptic inhibitory circuit. *Cell* *139*, 161–174.
- Fink, A.J., Croce, K.R., Huang, Z.J., Abbott, L.F., Jessell, T.M., and Azim, E. (2014). Presynaptic inhibition of spinal sensory feedback ensures smooth movement. *Nature* *509*, 43–48.
- Cazalets, J.R., Cournil, I., Geffard, M., and Moulins, M. (1987). Suppression of oscillatory activity in crustacean pyloric neurons: implication of GABAergic inputs. *J. Neurosci.* *7*, 2884–2893.
- Cinelli, E., Mutolo, D., Robertson, B., Grillner, S., Contini, M., Pantaleo, T., and Bongianini, F. (2014). GABAergic and glycinergic inputs modulate rhythmic mechanisms in the lamprey respiratory network. *J. Physiol.* *592*, 1823–1838.
- Schmitt, D.E., Hill, R.H., and Grillner, S. (2004). The spinal GABAergic system is a strong modulator of burst frequency in the lamprey locomotor network. *J. Neurophysiol.* *92*, 2357–2367.
- Cazalets, J.R., Bertrand, S., Sqalli-Houssaini, Y., and Clarac, F. (1998). GABAergic control of spinal locomotor networks in the neonatal rat. *Ann. N Y Acad. Sci.* *860*, 168–180.
- Kolmer, W. (1921). Das “Sagittalorgan” der Wirbeltiere. *Z. Anat. Entwicklungs* *60*, 652–717.
- Kolmer, W. (1931). Über das Sagittalorgan, ein Zentrales Sinnesorgan der Wirbeltiere, Insbesondere Beim Affen. *Z. Zellforsch. Mik. Ana.* *13*, 236–248.
- Agduhr, E. (1922). Über ein Zentrales Sinnesorgan (?) bei den Vertebraten. *Z. Anat. Entwicklungs* *66*, 223–360.
- Stoeckel, M.E., Uhl-Bronner, S., Hugel, S., Veinante, P., Klein, M.J., Mutterer, J., Freund-Mercier, M.J., and Schlichter, R. (2003). Cerebrospinal fluid-contacting neurons in the rat spinal cord, a gamma-aminobutyric acid system expressing the P2X2 subunit of purinergic receptors, PSA-NCAM, and GAP-43 immunoreactivities: light and electron microscopic study. *J. Comp. Neurol.* *457*, 159–174.
- Dale, N., Roberts, A., Ottersen, O.P., and Storm-Mathisen, J. (1987). The morphology and distribution of “Kolmer-Agduhr cells”, a class of cerebrospinal-fluid-contacting neurons revealed in the frog embryo spinal cord by GABA immunocytochemistry. *Proc. R. Soc. Lond. B Biol. Sci.* *232*, 193–203.
- Wyart, C., Del Bene, F., Warp, E., Scott, E.K., Trauner, D., Baier, H., and Isacoff, E.Y. (2009). Optogenetic dissection of a behavioural module in the vertebrate spinal cord. *Nature* *461*, 407–410.
- Lanuz, G.M., Gosgnach, S., Pierani, A., Jessell, T.M., and Goulding, M. (2004). Genetic identification of spinal interneurons that coordinate left-right locomotor activity necessary for walking movements. *Neuron* *42*, 375–386.
- Kiehn, O., Quinlan, K.A., Restrepo, C.E., Lundfald, L., Borgius, L., Talpalar, A.E., and Endo, T. (2008). Excitatory components of the mammalian locomotor CPG. *Brain Res. Brain Res. Rev.* *57*, 56–63.
- McLean, D.L., Masino, M.A., Koh, I.Y., Lindquist, W.B., and Fetcho, J.R. (2008). Continuous shifts in the active set of spinal interneurons during changes in locomotor speed. *Nat. Neurosci.* *11*, 1419–1429.
- Ritter, D.A., Bhatt, D.H., and Fetcho, J.R. (2001). In vivo imaging of zebrafish reveals differences in the spinal networks for escape and swimming movements. *J. Neurosci.* *21*, 8956–8965.
- McLean, D.L., Fan, J., Higashijima, S., Hale, M.E., and Fetcho, J.R. (2007). A topographic map of recruitment in spinal cord. *Nature* *446*, 71–75.
- Huang, A.L., Chen, X., Hoon, M.A., Chandrashekar, J., Guo, W., Tränkner, D., Ryba, N.J., and Zuker, C.S. (2006). The cells and logic for mammalian sour taste detection. *Nature* *442*, 934–938.
- Djenoune, L., Khabou, H., Joubert, F., Quan, F.B., Nunes Figueiredo, S., Bodineau, L., Del Bene, F., Burcklé, C., Tostivint, H., and Wyart, C. (2014). Investigation of spinal cerebrospinal fluid-contacting neurons expressing PKD2L1: evidence for a conserved system from fish to primates. *Front. Neuroanat.* *8*, 26.
- Orts-Del’Imagine, A., Kastner, A., Tillement, V., Tardivel, C., Trouslard, J., and Wanaverbecq, N. (2014). Morphology, distribution and phenotype of polycystin kidney disease 2-like 1-positive cerebrospinal fluid contacting neurons in the brainstem of adult mice. *PLoS ONE* *9*, e87748.
- Vigh, B., and Vigh-Teichmann, I. (1973). Comparative ultrastructure of the cerebrospinal fluid-contacting neurons. *Int. Rev. Cytol.* *35*, 189–251.
- Christenson, J., Alford, S., Grillner, S., and Hökfelt, T. (1991). Co-localized GABA and somatostatin use different ionic mechanisms to hyperpolarize target neurons in the lamprey spinal cord. *Neurosci. Lett.* *134*, 93–97.
- Meyer, M.P., and Smith, S.J. (2006). Evidence from in vivo imaging that synaptogenesis guides the growth and branching of axonal arbors by two distinct mechanisms. *J. Neurosci.* *26*, 3604–3614.
- Hale, M.E., Ritter, D.A., and Fetcho, J.R. (2001). A confocal study of spinal interneurons in living larval zebrafish. *J. Comp. Neurol.* *437*, 1–16.
- Satou, C., Kimura, Y., and Higashijima, S. (2012). Generation of multiple classes of V0 neurons in zebrafish spinal cord: progenitor heterogeneity and temporal control of neuronal diversity. *J. Neurosci.* *32*, 1771–1783.
- Kinkhabwala, A., Riley, M., Koyama, M., Monen, J., Satou, C., Kimura, Y., Higashijima, S., and Fetcho, J. (2011). A structural and functional ground plan for neurons in the hindbrain of zebrafish. *Proc. Natl. Acad. Sci. USA* *108*, 1164–1169.
- Koyama, M., Kinkhabwala, A., Satou, C., Higashijima, S., and Fetcho, J. (2011). Mapping a sensory-motor network onto a structural and functional ground plan in the hindbrain. *Proc. Natl. Acad. Sci. USA* *108*, 1170–1175.
- Higashijima, S., Mandel, G., and Fetcho, J.R. (2004). Distribution of prospective glutamatergic, glycinergic, and GABAergic neurons in embryonic and larval zebrafish. *J. Comp. Neurol.* *480*, 1–18.
- Higashijima, S., Schaefer, M., and Fetcho, J.R. (2004). Neurotransmitter properties of spinal interneurons in embryonic and larval zebrafish. *J. Comp. Neurol.* *480*, 19–37.
- Roy, B., and Ali, D.W. (2014). Multiple types of GABA responses identified from zebrafish Mauthner cells. *Neuroreport* *25*, 1232–1236.
- Grillner, S., McClellan, A., Sigvardt, K., Wallén, P., and Wilén, M. (1981). Activation of NMDA-receptors elicits “fictive locomotion” in lamprey spinal cord in vitro. *Acta Physiol. Scand.* *113*, 549–551.
- McDearmid, J.R., and Drapeau, P. (2006). Rhythmic motor activity evoked by NMDA in the spinal zebrafish larva. *J. Neurophysiol.* *95*, 401–417.
- Talpalar, A.E., Bouvier, J., Borgius, L., Fortin, G., Pierani, A., and Kiehn, O. (2013). Dual-mode operation of neuronal networks involved in left-right alternation. *Nature* *500*, 85–88.
- Bellardita, C., and Kiehn, O. (2015). Phenotypic characterization of speed-associated gait changes in mice reveals modular organization of

- locomotor networks. *Curr. Biol.* 25, 1426–1436, <http://dx.doi.org/10.1016/j.cub.2015.04.005>.
37. Shik, M.L., Severin, F.V., and Orlovsky, G.N. (1969). Control of walking and running by means of electrical stimulation of the mesencephalon. *Electroencephalogr. Clin. Neurophysiol.* 26, 549.
 38. Dubuc, R., Brocard, F., Antri, M., Fénelon, K., Gariépy, J.F., Smetana, R., Ménard, A., Le Ray, D., Viana Di Prisco, G., Pearlstein, E., et al. (2008). Initiation of locomotion in lampreys. *Brain Res. Brain Res. Rev.* 57, 172–182.
 39. Quan, F.B., Dubessy, C., Galant, S., Kenigfest, N.B., Djenoune, L., Leprince, J., Wyart, C., Lihmann, I., and Tostivint, H. (2015). Comparative distribution and in vitro activities of the urotensin II-related peptides URP1 and URP2 in zebrafish: evidence for their colocalization in spinal cerebrospinal fluid-contacting neurons. *PLoS ONE* 10, e0119290.
 40. Jean-Xavier, C., Mentis, G.Z., O'Donovan, M.J., Cattaert, D., and Vinay, L. (2007). Dual personality of GABA/glycine-mediated depolarizations in immature spinal cord. *Proc. Natl. Acad. Sci. USA* 104, 11477–11482.
 41. Marder, E., and Bucher, D. (2001). Central pattern generators and the control of rhythmic movements. *Curr. Biol.* 11, R986–R996.
 42. Roberts, A., and Tunstall, M.J. (1990). Mutual re-excitation with post-inhibitory rebound: a simulation study on the mechanisms for locomotor rhythm generation in the spinal cord of *Xenopus* embryos. *Eur. J. Neurosci.* 2, 11–23.
 43. Soffe, S.R. (1990). Active and passive membrane properties of spinal cord neurons that are rhythmically active during swimming in *Xenopus* embryos. *Eur. J. Neurosci.* 2, 1–10.
 44. Roberts, A., Li, W.C., and Soffe, S.R. (2008). Roles for inhibition: studies on networks controlling swimming in young frog tadpoles. *J. Comp. Physiol. A Neuroethol. Sens. Neural Behav. Physiol.* 194, 185–193.
 45. Pirtle, T.J., and Satterlie, R.A. (2007). The role of postinhibitory rebound in the locomotor central-pattern generator of *Clione limacina*. *Integr. Comp. Biol.* 47, 451–456.
 46. Bertrand, S., and Cazalets, J.R. (1998). Postinhibitory rebound during locomotor-like activity in neonatal rat motoneurons in vitro. *J. Neurophysiol.* 79, 342–351.
 47. Orts-Del'immagine, A., Wanaverbecq, N., Tardivel, C., Tillement, V., Dallaporta, M., and Trouslard, J. (2012). Properties of subependymal cerebrospinal fluid contacting neurones in the dorsal vagal complex of the mouse brainstem. *J. Physiol.* 590, 3719–3741.
 48. Masino, M.A., and Fetcho, J.R. (2005). Fictive swimming motor patterns in wild type and mutant larval zebrafish. *J. Neurophysiol.* 93, 3177–3188.